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**Research** Article

# Multi-environment analysis of grain yield in a diverse set of bread wheat genotypes

Charan Singh\*, Arun Gupta, Pradeep Kumar, Sendhil Ramdas, Gopalareddy Krishanappa, Vikas Gupta, Sanjay Kumar Singh, Amit Kumar Sharma, Bhudeva Singh Tyagi, Gyanendra Singh, Ravish Chatrath and Gyanendra Pratap Singh

ICAR-Indian Institute of Wheat and Barley Research, Karnal, India - 132001

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#### \*Corresponding author

Email: n\_charansingh@hotmail.com

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#### 1. Introduction

Wheat is a staple food crop of many countries across the globe including India, which plays an important role in nutritional as well as food security. India is one of the prime producer and consumer of wheat. In India, wheat was cultivated over 29.55 million hectares with a production of 101.20 million tons during 2018-19 (3<sup>rd</sup> estimates, Anonymous, 2019). India is divided into five mega agro-climatic zones for wheat cultivation i.e., Northern Hill Zone, North Western Plains Zone, North Eastern Plains Zone, Central Zone and Peninsular Zone. Climate change and increasing human population around the globe are the most critical challenges to meet the ever increasing demand of food grain production for the growing population. Recently, major wheat growing regions are experiencing extreme and unpredictable weather conditions due to climate change. Therefore, the development of high yielding climate resilient wheat genotypes is of paramount importance to meet the global demand of wheat production. Due to these challenges,

Abstract

A diverse set of 50 bread wheat genotypes was evaluated at 16 different locations belonging to four major wheat growing zones of India during 2016-17. The yield data was analyzed using Eberhart and Russel (1966), AMMI and GGE biplot stability models. Genotypes G14 and G26 were identified as the most stable genotypes based on Eberhart and Russel (1966) model, whereas genotypes G10, G14, G18, G23, G26, G30, and G47 were identified stable as per AMMI model whereas five genotypes G26, G23, G10, G43 and G14 were found stable as per GGE biplot analysis. Genotypes G14 and G26 were turned out to be the most stable genotypes based on these three stability models. The G48 ranked best for (Durgapura and Pantnagar); G29 (Powarkheda) and G35 (Hisar, Delhi and Ludhiana) based on grain yield performance. The GGE bi-plot indicated Durgapura to be an ideal environment followed by Hisar, Ludhiana, Delhi, Pantnagar, and Powarkheda that could be recommended for the future wheat breeding programmes aiming towards yield enhancement.

**Keywords:** AMMI, GGE, genotype × environment interaction, gain yield stability, wheat

wheat production needs to be sustainably increased by 2-3% annually. Gene expression is subjected to modification by the environment; therefore, genotypic expression of the phenotypes is environmentally dependent (Kang, 1998). The performance of genotypes is greatly affected by genotype-environment interaction (GEI)(Becker and Leon, 1988). The major focus of plant breeding community across the globe is to develop genotypes either with general adaptability and/or specific adaptability (Ebdon and Gauch, 2002a). The complexity of GEI makes selection procedure cumbersome to identify the best performing and most stable genotypes (Yau, 1995). The adaptability of a variety over diverse environments is usually tested by the degree of its interaction with different environmental factors under which it is grown. Stability, or genotype (G) x environment (E) interactions, is necessary for researcher's to develop a genotype that respond optimally and consistently across environments (Blanchi et al. 2008). A variety or genotype is considered to be more adaptive or stable one, if it has a high mean yield but low degree of fluctuation in yielding ability,

when grown over diverse environments. GE interactions exist when the responses of two genotypes to different levels of environmental stress are crossovers (Allard and Bradshaw, 1964). Numerous tools have been developed to measure the response of genotypes to changes in environment (Eberhart and Russell, 1966; Shukla, 1972; Gauch, 1988; Lin and Binns, 1988). Eberhart and Russell (1966) (ER) regression model describes a stable variety which should have around unit regression coefficient over environments (bi=1) and minimum deviation from the regression (S<sup>2</sup>di=0) in addition to higher grain yield than population mean. The other popular stability models additive main effect and multiplicative interaction (AMMI) and genotype plus genotype-by-environment (GGE) overcomes the limitations of univariate models and also are more informative compared to other methods (Shukla, 1972; Gauch, 1988; Zobel et al. 1988). However, GGE biplot offers breeders a more complete and visual evaluation of all aspects of the data by creating a biplot that simultaneously represents both mean performance and stability (Yan, 2001). Widespread acceptance of GGE biplot for its ability to evaluate mean performance and stability and to identify mega-environments has created a need for research to compare GGE Biplot to other "traditional" stability analysis tools.

AMMI and *GGE* are the two commonly used bi-plots techniques to visualize G×E interactions (Yan *et al.* 2000). The GGE biplot is an effective method based on principal component analysis (PCA) to fully explore multienvironmental data (Rao *et al.* 2011; Singh *et al.* 2019). It has always been challenging to define how new genotypes would respond under different climatic conditions, without graphically presenting the data, when many cultivars are evaluated across many sites, seasons and years (Yan *et al.*  2001). The researchers usually focus on  $G \times E$  interaction to select the genotypes for general adaptation and crossover  $G \times E$  interaction for the specific adaptation (Matus-Cadiz *et al.* 2003).

Development of high buffering genotypes which adapts to diverse agro-climatic conditions is both economical and sustainable strategy to sustain the wheat production in changing climatic conditions. In this study, genotypes received from CIMMYT as a Semi-Arid Wheat Yield Trial (SAWYT) were evaluated for grain yield across different environments to identify stable genotypes for general and specific adaptation. Therefore, the present study was aimed (i) to identify mega-environments in major wheat growing regions of India (ii) to identify high yielding and stable performing genotypes.

#### 2. Materials and methods

A set of 50 advanced generation bread wheat genotypes of 24th Semi-Arid Wheat Yield Trial (24th SAWYT) of CIMMYT were grown at16 locations in India during rabi, 2016-17 (Table 1 and 2). The genotypes were planted in an randomised complete block design with two replications per genotype in a six row plot of 6 m length with a plant-to-plant distance and row to row distance of 10cm and 25cm, respectively. Grain yield from 4 middle rows with a net plot size of 4.8 m<sup>2</sup> was converted into q ha-1 at 10% standard grain moisture and utilized for further statistical analysis. Standard agronomic practices were followed for raising a healthy crop. The data of 50 genotypes were subjected to pooled analysis of variance using SAS statistical package. A principal component analysis (PCA) was performed to construct a GGE and AMMI biplot using IRRI-STAT software (ver. 5.0). The stability parameters for grain yield were worked out as

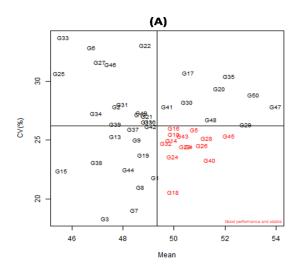


Fig 1 Bi plot of CV for grain yield

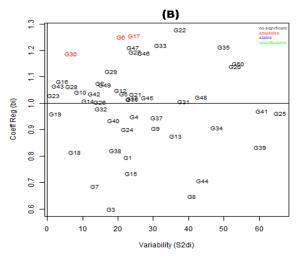


Fig 2 Bi plot of stability parameters for grain yield

**Table1.** Details of 16 locations (environments) used forconduction of experiments.

Env. code	Location	Latitude	Longitude	Mean sea level (m)	Mean grain yield (qha <sup>-1</sup> )
E1	Delhi	28º39'N	77º13'E	227.0	46.51
E2	Karnal	29º41'N	76°59'E	253.0	71.93
E3	Pantnagar	29º30'N	79º31'E	243.8	60.14
E4	Hisar	29º10'N	75º 46'E	215.2	55.07
E5	Ludhiana	30°54'N	75°51'E	252.0	60.37
E6	Durgapura	26°55'N	75°46'E	432.0	57.96
E7	Faizabad	26º46'N	82º8'E	104.0	38.36
E8	Kota	25°10'N	75°50'E	267.0	65.81
E9	Powarkheda	21º50'N	76º43'E	229.0	74.08
E10	Vijapur	21º11'N	72°49E	20.0	36.76
E11	Junagadh	21º52'N	70º46'E	97.0	54.42
E12	Jabalpur	23°10'N	79°57'E	416.0	39.00
E13	Pune	18º31'N	73°51'E	562.0	60.10
E14	Indore	22º43'N	75°51'E	550.0	48.89
E15	Bilaspur	22º4'N	82°9'E	264.0	33.67
E16	Dharwad	15°27'N	75°0'E	724.0	35.09

per procedure (Eberhart and Russell 1966; Singh and Chaudhary, 1985).

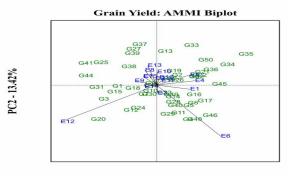




Fig 3 Bi plot depicting the relationship among test environments by AMMI

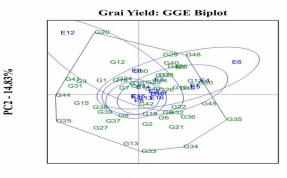
#### 3. Results and discussion

#### 3.1 Genetic variability

The genotypes under study showed a wide spectrum of variability with respect to their mean value of grain yield at different locations (Table 1). The location wise mean yield ranges from 33.67q/ha (Bilaspur) to 74.08 q/ha (Powarkheda). The highest yielding genotypes at individual test locations are G5 (63.82q/ha) at Delhi, G47 (55.25q/ha) at Karnal, G11 (50.25q/ha) at Pantnagar, G50 (78.96q/ha) at Hisar, G50 (70.67q/ha) at Ludhiana, G48 (73.44q/ha) at Durgapura, G44 (47.40q/ha) at Faizabad, G21 (76.98q/ha) at Kota, G20 (86.30q/ha) at Powarkheda, G47 (52.92q/ha) at Vijapur, G29 (68.87q/ha) at Junagadh, G20 (63.37q/ha) at Jabalpur, G26 (68.47q/ha) at Pune, G10 (48.60q/ha) at Indore, G18 (39.10)q/ha at Bilaspur and G39 (49.35q/ha) at Dharwad. Out of 50 genotypes, only two genotypes (G50 and G47) ranked first at two locations each. Genotype G50 was the top yielder at Hisar and Ludhiana whereas G47 at Karnal and Vijapur locations. The differential ranking of genotypes showed their crossover type of GEI effects. This differential expression of genotypes with respect to gain yield is due to greater magnitude of environmental effects in their complete expression. The coefficient of variation (CV) of the genotypes ranged from 18.31% (G3) to 33.69% (G33). The genotypes with high mean values with comparatively low CV are G5, G10, G14, G16, G18, G24, G26, G28, G32, G43, G45 and G46 are considered to be desirable (Table 4 and Fig.1).

#### 3.2. Stability analysis by Eberhart and Russell model

The genotype-environment interaction component (GEI) was further elaborated by using the joint regression model of stability analysis (Eberhart and Russell, 1966). Stable genotypes were identified based on stability parameters viz, linear response (bi=1), minimum deviation from linearity (S<sup>2</sup>di= 0), and higher trait mean. The genotypes



PC1 - 23.24%

Fig 4 Bi plot depicting the relationship among environments by GGE

*viz*, G5, G10, G16, G17, G20, G23, G28, G29, G30, G35, G43, G45, G47, G48 and G50 were specifically suited to high productive/favorable environments due to their higher mean values than population mean, regression coefficient  $(b_i)$  more than unity with non-significant deviations from their respective regression. Similarly, the genotypes G4, G18, G24, G32, G41, G40, G125 and G126are better suited to low productive/unfavorable environments due to their higher mean values than populations mean coupled with regression coefficient

Table 2. Pedigree	details of 50	genotypes use	ed in the	present study

Code	Pedigree details
G1	LOCAL CHECK
G2	PBW65/2*PASTOR
G3	KA/NAC//TRCH/3/DANPHE #1
G4	FITIS
G5	FRANCOLIN #1//WBLL1*2/BRAMBLING
G6	FRANCOLIN #1/3/PBW343*2/KUKUNA*2//YANAC
G7	BECARD #1/3/PBW343*2/KUKUNA//PBW343*2/KUKUNA
G8	SUP152/3/TRCH/SRTU//KACHU
G9	WAXWING/7/TNMU/6/CEP80111/CEP81165/5/IAC5/4/YKT406/3/AG/ASN//ATR/8/ ATTILA/3*BCN//BAV92/3/TILHI/4/SHA7/VEE#5//ARIV92
G10	KACHU/SAUAL/3/TRCH/SRTU//KACHU
G11	ROLF07/4/WBLL1/KUKUNA//TACUPETO F2001/3/UP2338*2/VIVITSI/5/SAUAL/MUTUS
G12	WHEAR/KUKUNA/3/C80.1/3*BATAVIA//2*WBLL1*2/6/WBLL1*2/4/YACO/PBW65/3/ KAUZ*2/TRAP//KAUZ/5/KACHU #1
G13	PBW343*2/KUKUNA//PBW343*2/KUKUNA*2/8/SHA7//PRL/VEE#6/3/FASAN/4/ HAAS8446/2*FASAN/5/CBRD/KAUZ/6/MILAN/AMSEL/7/FRET2*2/KUKUNA
G14	PRL/2*PASTOR*2/5/CROC_1/AE.SQUARROSA (205)//BORL95/3/PRL/SARA//TSI/ VEE#5/4/FRET2
G15	KACHU*2/6/YAR/AE.SQUARROSA (783)/4/GOV/AZ//MUS/3/SARA/5/MYNA/VUL//JUN
G16	HEILO//MILAN/MUNIA/3/2*WHEAR/SOKOLL
G17	WAXWING//INQALAB 91*2/KUKUNA/3/WBLL1*2/TUKURU/8/2*NG8201/KAUZ/4/ SHA7//PRL/VEE#6/3/FASAN/5/ MILAN/KAUZ/6/ACHYUTA/7/PBW343*2/KUKUNA
G18	HUW234+LR34/PRINIA*2//SNLG/3/BOKOTA
G19	CHEN/AE.SQ//WEAVER/3/SSERI1/4/TOBA97/PASTOR/5/MUU #1/6/KACHU #1//PI 610750/SASIA/3/KACHU
G20	COPIO/5/UP2338*2/SHAMA/3/MILAN/KAUZ//CHIL/CHUM18/4/UP2338*2/SHAMA
G21	SAUAL/YANAC//SAUAL/5/UP2338*2/SHAMA/3/MILAN/KAUZ//CHIL/CHUM18/4/ UP2338*2/SHAMA
G22	PRL/2*PASTOR/3/2*TRCH/SRTU//KACHU
G23	FRANCOLIN #1/3/PBW343*2/KUKUNA*2//YANAC/4/KINGBIRD #1//INQALAB 91*2/ TUKURU
G24	PRL/2*PASTOR//SUNSTATE/4/2*ATTILA*2/PBW65//PIHA/3/ATTILA/2*PASTOR
G25	BAV92//IRENA/KAUZ/3/HUITES/4/DOLL*2/5/FRET2/KUKUNA//FRET2/3/TNMU/4/ FRET2*2/SHAMA
G26	TACUPETO F2001/BRAMBLING/5/NAC/TH.AC//3*PVN/3/MIRLO/BUC/4/2*PASTOR*2/6/ TRCH/SRTU//KACHU
G27	KAUZ//ALTAR 84/AOS/3/MILAN/KAUZ/4/SAUAL/5/PBW343*2/KUKUNA//PARUS/3/ PBW343*2/KUKUNA/6/KACHU/SAUAL
G28	SAUAL/MUTUS//KINGBIRD #1/3/SAUAL/MUTUS
G29	SAUAL/MUTUS*2/3/TRCH/SRTU//KACHU
G30	SAUAL/MUTUS*2//PICAFLOR #1
G31	C80.1/3*BATAVIA//2*WBLL1/5/REH/HARE//2*BCN/3/CROC_1/AE.SQUARROSA (213)// PGO/4/HUITES*2/6/TRCH/SRTU// KACHU
G32	ROLF07*2/KIRITATI*2//PICAFLOR #1
G33	FRET2*2/SHAMA//PARUS/3/FRET2*2/KUKUNA*2/4/TRCH/SRTU//KACHU
G34	FRET2*2/SHAMA//PARUS/3/FRET2*2/KUKUNA*2/4/TRCH/SRTU//KACHU

- G35 ATTILA\*2/PBW65\*2//WHEAR/3/TRCH/SRTU//KACHU/8/ALD/CEP75630//CEP75234/ PT7219/3/BUC/BJY/4/CBRD/5/TNMU/PF85487/6/PBW343\*2/KUKUNA/7/CNO79// PF70354/MUS/3/PASTOR/4/BAV92
- G36 BCN/RIALTO//2\*MUNAL #1
- G37 CROC\_1/AE.SQUARROSA(205)//BORL95/3/PRL/SARA//TSI/VEE#5/4/FRET2/6/ MTRWA92.161/PRINIA/5/SERI\*3//RL6010/4\*YR/3/PASTOR/4/BAV92
- G38 BAVIS/8/BOW/VEE/5/ND/VG9144//KAL/BB/3/YACO/4/CHIL/6/CASKOR/3/CROC\_1/ AE.SQUARROSA (224)//OPATA/7/ PASTOR//MILAN/KAUZ/3/BAV92
- G39 SOKOLL/3/PASTOR//HXL7573/2\*BAU/4/SHAMA//PARUS/PASTOR
- G40 W15.92/4/PASTOR//HXL7573/2\*BAU/3/WBLL1/8/BOW/VEE/5/ND/VG9144//KAL/BB/3/ YACO/4/CHIL/6/CASKOR/3/CROC\_1/AE.SQUARROSA (224)//OPATA/7/PASTOR// MILAN/KAUZ/3/BAV92
- G41 BECARD #1/BAVIS
- G42 WBLL1\*2/BRAMBLING/3/SOKOLL//SUNCO/2\*PASTOR
- G43 PASTOR//HXL7573/2\*BAU/3/WBLL1/6/MTRWA92.161/PRINIA/5/SERI\*3//RL6010/4\*YR/3/ PASTOR/4/BAV92
- G44 OASIS/5\*BORL95/5/CNDO/R143//ENTE/MEXI75/3/AE.SQ/4/2\*OCI/6/SOKOLL// SUNCO/2\*PASTOR/7/SOKOLL//SUNCO/2\*PASTOR
- G45 SOKOLL/3/PASTOR//HXL7573/2\*BAU\*2/6/OASIS/5\*BORL95/5/CNDO/R143//ENTE/ MEXI75/3/AE.SQ/4/2\*OCI
- G46 GLADIUS/5/2\*W15.92/4/PASTOR//HXL7573/2\*BAU/3/WBLL1
- G47 CROC\_1/AE.SQUARROSA (205)//BORL95/3/PRL/SARA//TSI/VEE#5/4/FRET2/5/2\*BAVIS #1
- G48 D67.2/PARANA 66.270//AE.SQUARROSA (320)/3/CUNNINGHAM/4/PASTOR/SLVS/5/ SUNCO/2\*PASTOR//EXCALIBUR/ 6/MTRWA92.161/PRINIA/5/SERI\*3//RL6010/4\*YR/3/ PASTOR/4/BAV92
- G49 LIVINGSTON/5/2\*W15.92/4/PASTOR//HXL7573/2\*BAU/3/WBLL1
- G50 MUNAL #1\*2//SOKOLL/WBLL1

Table	3.	ANOVA	for	AMMI	analysis	of	50	diverse
genoty	pes i	n 16 enviro	onm	ents.				

Source of variation	df	MS	F	Prob
Environments	15.00	13159.79	549.15	< 0.00
Genotypes	49.00	120.81	5.04	< 0.00
G x E	735.00	78.60	3.28	< 0.00
PC1	63.00	219.75	9.21	< 0.00
PC2	61.00	127.06	5.32	< 0.00
PC3	59.00	120.43	5.05	< 0.00
PC4	57.00	109.51	4.59	< 0.00
PC5	55.00	89.77	3.76	< 0.00
PC6	53.00	73.10	3.06	< 0.00
PC7	51.00	56.43	2.36	< 0.00
PC8	49.00	48.82	2.05	< 0.00
PC9	47.00	44.36	1.86	< 0.00
PC10	45.00	38.07	1.59	< 0.00
Residuals	800.00	23.96	NA	NA

 $(b_i)$  less than unity with non-significant deviations from regression. Genotypes G14 and G26 exhibited stable performance across the environments suggested by their higher mean performance than population mean, regression coefficient ( $b_i$ ) near to unity with non-significant deviations from regression. Kumar *et al.* (2017) identified genotype K 612 to be stable as it had superior mean performance, regression coefficient greater than unity with non-significant deviation from regression coefficient. Another genotype K 9162 was the highest yielding and stable genotype along with early maturity (118 days) and has potential for late planting conditions.

#### 3.3. Stability analysis by AMMI model

The AMMI model is widely used in the stability analysis as it provides an initial diagnosis of the model to be fit into multi- environmental evaluation (Zobel *et al.* 1988, Crossa *et al.* 1990). Analysis of variance (ANOVA) revealed significant role of the additive main effects *i.e.*, genotype and environment on the total variation of the traits (Table 3). The multiplicative analysis revealed that, all the 10 interaction principal components were significant. The first two interaction principal components cumulatively explained 37.38% of interaction. Graphical representation (biplot) employed by using PCA1 and PCA2 to ascertain environmental variation and to interpret the genotype-environment interactions (Fig.3).

**Table 4.** Stability parameters (Eberhart and Russell model for grain yield of 50 genotypes in 16 locations

ioi grain yie	iu or or g	senotype	s III 10 10Ca	uions	
Genotype	Mean	SD	CV (%)	bi	S <sup>2</sup> di
G1	49.28	10.75	21.81	0.80	22.91
G2	47.74	13.29	27.84	1.07	14.85
G3	47.29	8.66	18.31	0.60	18.13
G4	50.57	12.35	24.41	0.95	24.58
G5	50.79	13.13	25.84	1.04	21.62
G6	46.75	15.36	32.86	1.25	20.82
G7	48.46	9.23	19.05	0.69	13.36
G8	48.67	10.21	20.98	0.65	40.87
G9	48.54	12.13	24.99	0.90	30.73
G10	49.99	12.75	25.50	1.04	9.30
G11	48.94	12.99	26.55	1.02	23.90
G12	48.69	13.21	27.14	1.05	20.70
G13	47.69	12.07	25.30	0.88	36.35
G14	49.91	12.46	24.97	1.01	11.52
G15	45.58	10.20	22.37	0.73	23.76
G16	49.99	13.00	26.00	1.08	4.19
G17	50.56	15.53	30.72	1.26	24.60
G18	49.97	10.27	20.55	0.81	7.75
G19	48.80	11.58	23.73	0.96	2.25
G20	51.77	15.21	29.37	1.14	53.20
G21	48.95	13.22	27.00	1.03	25.02
G22	48.87	16.14	33.03	1.28	37.56
G23	50.45	12.32	24.43	1.03	1.74
G24	49.94	11.78	23.59	0.90	22.65
G25	45.49	13.94	30.64	0.96	65.90
G26	51.10	12.55	24.55	1.00	14.81
G27	47.08	14.88	31.60	1.19	24.79
G28	51.28	12.89	25.13	1.06	6.96
G29	52.81	13.89	26.31	1.12	18.06
G30	50.52	14.24	28.19	1.19	6.66
G31	47.97	13.43	27.99	1.01	38.65
G32	49.70	12.29	24.73	0.98	15.26
G33	45.66	15.38	33.69	1.22	32.02
G34	46.95	12.80	27.26	0.91	47.97
G35	52.14	15.85	30.39	1.21	49.95
G36	49.05	13.01	26.52	1.02	24.11
G37	48.39	12.55	25.93	0.95	30.96
G38	46.97	10.84	23.09	0.82	19.28
G39	47.71	12.58	26.37	0.83	60.24
G40	51.41	11.96	23.27	0.93	18.68

G41	49.75	13.85	27.83	0.97	60.83
G42	49.07	12.83	26.14	1.04	13.28
G43	50.35	12.76	25.34	1.06	2.97
G44	48.20	10.84	22.49	0.71	43.87
G45	52.16	13.21	25.32	1.02	28.31
G46	47.50	14.91	31.39	1.19	27.32
G47	53.98	15.03	27.85	1.21	24.19
G48	51.46	13.76	26.74	1.02	43.56
G49	48.72	13.30	27.29	1.07	16.36
G50	53.12	15.33	28.85	1.15	54.00

Genotypes with PCA1 scores near zero showed very less interaction across environments. Similarly, environments with PCA1 scores near zero displaying little interaction across genotypes and generally show less discrimination among the genotypes (Crossa *et al.* 1990). Jabalpur and Durgapura are the distinct environments compared to other locations (Fig.3).

The genotypes *i.e.*, G10, G14, G18, G23, G26, G30 and G47 showed greater stability since they found very close to the origin (Fig.3). The genotypes which have the longest vectors were adapted to specific environments and contributed to more diversity. The genotypes appearing close together on the plot have almost similar mean grain yield over all the locations while the genotypes placed far apart may either differ in mean grain yield or show a different pattern of response over the locations. Hence, the genotypes near to the origin are not sensitive to genotype × environment interactions whereas those distant from the origins are more sensitive and exhibit large genotype × environment interactions. The genotypes located near the centre of the bi-plot contributed less to GE interaction, while cultivars having longer vectors, showed the greatest contribution to GE interaction (Letta et al. 2008). The distance from the origin exhibits the amount of interaction by genotypes either over environments or by environments over genotypes (Yan and Kang 2003). The genotypes viz, G20, G33, G35, G37, G41, G44, G46 and G48 expressed interaction on higher side (positively or negatively). The genotypes at vertex were the winners in the environment included in that sector (Yan and Tinker, 2006). The genotypes that results in both high mean grain yield and high stability termed as an ideal genotype, should possess both high mean performance and high stability across environments. The environment with short spike (Pantnagar, Vijapur, Hisar and Junagadh) does not exert strong interactive forces. While the environment with long spike (Jabalpur, Powarkheda, Durgapura, Ludhiana) exert strong interaction among each other.

#### Multi-environment analysis of grain yield in a diverse bread wheat genotypes

Table 5. Mean grain yield  $(q/ha^{-1})$  performance of 50 genotypes for grain yield across 16 locations

Entry	Delhi	Karnal	Pantnagar	Hisar	Ludhiana	Durgapura	Faizabad	Kota	Powarkheda	Vijapur	Junagadh	Jabalpur	Pune	Indore	Bilaspur	Dharwad
Gl	53.61	40.67	49.42	44.71	56.07	48.96	38.04	59.90	78.96	44.58	51.22	47.43	55.41	48.01	38.47	32.94
G2	43.89	45.26	43.48	55.88	50.82	58.33	41.67	74.27	68.75	26.50	56.33	29.08	55.81	37.89	35.28	40.64
G3	43.61	40.04	42.43	50.92	51.95	49.48	40.64	58.75	61.04	40.00	55.90	57.20	55.00	37.15	36.11	36.45
G4	53.54	42.99	50.18	54.21	66.53	71.88	32.00	63.02	65.63	32.42	55.35	44.27	56.16	43.94	35.83	41.24
G5	63.82	43.26	48.43	60.92	61.10	63.54	38.16	58.02	80.25	38.42	53.32	40.97	50.25	41.33	28.96	41.91
G6	58.26	38.74	34.88	63.29	50.50	55.21	37.98	68.44	74.37	31.83	55.87	28.00	54.44	42.58	25.56	28.05
G7	47.78	41.11	47.25	56.92	59.45	52.60	35.02	58.96	58.33	35.08	56.12	48.00	57.78	48.36	37.01	35.49
G8	53.33	48.53	37.13	50.13	44.72	72.40	45.59	62.08	54.17	42.33	52.62	33.53	59.16	43.63	36.04	43.35
G9	56.32	43.92	46.03	35.79	47.03	57.29	41.67	71.04	70.83	35.67	60.58	34.27	56.03	44.93	35.90	39.38
G10	48.96	48.10	42.48	50.00	61.53	60.94	39.37	70.00	72.92	33.08	47.25	43.70	65.19	48.60	34.38	33.40
G11	46.39	44.15	50.25	47.67	60.37	69.79	40.34	54.06	81.25	37.17	50.65	37.77	53.25	41.79	31.53	36.64
G12	44.17	38.08	46.80	52.04	56.35	59.90	41.36	61.67	84.47	33.00	49.57	49.87	52.19	42.40	29.86	37.29
G13	40.13	45.68	44.85	43.33	62.15	48.44	36.53	73.33	62.08	37.50	60.08	28.50	56.81	40.98	35.76	46.81
G14	50.42	43.14	37.63	57.25	56.70	63.02	37.56	56.67	81.15	38.25	61.60	39.97	52.47	43.10	34.38	45.31
G15	36.88	46.90	40.10	43.21	45.30	50.00	30.50	55.73	67.71	36.17	59.92	47.03	54.59	38.03	32.51	44.71
G16	50.83	43.51	48.38	57.79	59.58	66.67	41.06	61.35	79.17	35.33	52.17	33.77	56.88	37.21	35.00	41.10
G17	52.78	47.76	49.98	64.58	50.07	71.35	33.82	62.81	85.05	31.17	53.35	31.03	60.97	38.94	34.86	40.49
G18	47.85	42.00	48.10	51.21	61.95	53.13	33.94	67.81	68.13	41.92	55.90	50.10	57.09	42.34	39.10	38.91
G19	45.83	46.56	43.88	53.08	58.82	56.77	46.68	65.42	71.67	38.83	54.13	32.73	58.44	34.00	34.79	39.14
G20	46.46	48.92	40.73	56.58	50.23	63.54	36.23	72.60	86.30	29.25	52.73	63.37	62.50	46.83	36.53	35.51
G21	56.25	49.89	37.00	55.58	55.33	60.42	37.98	76.98	62.08	33.92	56.80	27.65	55.94	42.33	33.47	41.52
G22	47.08	46.42	28.83	71.50	60.67	60.94	41.67	69.58	75.00	24.25	56.95	30.88	57.50	41.29	36.60	32.79
G23	51.74	45.81	41.78	61.96	57.32	60.94	39.25	72.40	70.00	30.75	53.00	44.00	59.19	43.40	34.65	40.99
G24	42.57	50.06	46.13	51.58	43.05	66.67	38.04	72.50	70.83	35.08	53.77	46.55	58.84	44.80	36.53	42.00
G25	46.11	42.44	41.05	51.04	31.58	38.02	30.19	72.19	80.21	31.75	48.57	40.10	55.97	40.03	37.15	41.49
G26	49.86	46.88	49.93	59.54	65.52	58.33	31.40	65.46	68.75	32.92	54.62	39.43	68.47	46.90	34.79	44.85
G27	39.38	43.36	39.93	43.67	55.68	44.79	38.35	71.88	84.54	34.67	52.23	31.58	62.59	40.05	32.08	38.44
G28	58.47	44.75	41.38	58.54	52.18	65.63	39.25	68.54	75.00	35.42	61.27	43.27	57.69	43.95	28.96	46.19
G29	51.74	44.78	45.78	61.17	56.27	70.83	41.55	72.71	77.29	37.42	68.87	47.77	53.09	46.84	31.46	37.36
G30	54.10	49.83	41.78	52.79	62.00	60.42	40.22	70.42	80.42	30.75	50.78	46.02	62.00	43.76	33.61	29.39
G31	45.56	46.96	41.40	46.00	47.55	49.48	42.27	75.73	78.13	31.08	49.10	50.48	56.31	43.48	36.60	27.38
G32	52.15	44.24	36.68	50.92	64.37	63.54	39.86	68.65	65.83	43.92	54.38	32.25	60.81	41.75	30.42	45.41
G33	48.61	47.43	33.95	45.58	59.43	48.96	37.44	63.33	83.33	32.25	50.32	16.87	54.72	34.70	34.44	39.21
G34	49.31	48.43	41.45	54.00	64.72	59.38	38.41	58.85	56.25	33.08	54.73	18.33	61.59	42.75	36.88	33.08
G35	61.81	46.32	44.13	70.67	68.70	61.46	34.54	72.29	70.83	42.17	57.23	20.13	64.88	42.38	35.28	41.46
G36	51.67	45.25	43.98	64.42	52.77	59.38	37.14	58.85	75.00	45.50	55.23	23.67	58.28	43.28	31.46	38.99
G37	48.26	54.29	49.40	55.25	54.32	37.50	38.16	65.10	78.34	38.92	51.13	33.27	58.63	42.36	33.89	35.44
G38	53.26	43.92	40.03	50.88	59.20	39.06	39.55	61.67	71.88	35.33	45.68	43.25	52.47	45.39	28.75	41.25
G39	44.03	42.14	32.85	45.00	66.82	42.19	40.34	54.48	79.17	50.75	51.03	41.08	57.97	35.70	30.42	49.35
G40	59.72	54.74	41.98	56.50	69.20	64.06	41.55	59.48	72.92	34.17	52.12	47.43	53.38	40.91	34.24	40.15
G41	49.51	49.13	41.38	49.75	63.90	36.98	41.97	65.63	80.42	31.42	51.33	56.57	67.09	38.68	34.03	38.20
G42	44.72	47.97	46.43	46.50	56.67	60.42	38.89	69.06	77.08	49.33	53.70	33.90	56.75	37.45	31.11	35.20
G42 G43	49.38	54.36	48.68	48.54	60.88	57.29	40.70	66.15	77.08	31.50	58.55	41.80	60.94	40.34	32.99	36.49
G43 G44	43.13	49.54	33.00	51.17	39.88	44.27	40.70	64.69	71.88	39.67	56.85	53.13	59.53	41.71	35.00	40.38
G44 G45	63.26	48.81	49.10	53.58	56.12	69.79	36.23	70.63	70.83	44.17	59.25	27.23	60.41	42.06	35.63	47.48
G45 G46	53.61	48.81	49.10	48.21	62.75	71.88	35.02	60.10	70.83	28.33	59.25	31.92	46.81	42.00 36.16	26.60	37.66
G40 G47	46.67	47.30 55.25	40.30	48.21 65.88			34.42		85.42	28.55 52.92		43.70	40.81 58.09	37.39	32.22	44.86
G47 G48					63.40	63.54 73.44		76.25			61.30 50.97					
	60.21 45.69	53.63 48.96	47.28	53.38 56.25	63.22 61.78	73.44	30.19	56.88 75.54	80.21	47.67		43.50	52.34 44.63	38.04	29.72	42.68
G49	45.69	48.96	43.83	56.25 78.06	61.78 70.67	61.46	37.44	75.54 58.54	72.71	36.58	48.35	40.43	44.63	35.48	31.18	39.19
G50	57.29 50.20	47.79 46.40	49.50 43.19	78.96 54.17	70.67 57.14	53.65 57.96	40.16 38.36	58.54 65.81	83.33 74.08	43.67 36.76	58.10 54.42	33.42 39.00	63.44 57.42	38.79 41.48	35.35 33.67	37.33 39.30

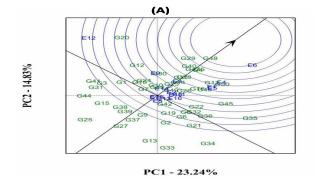


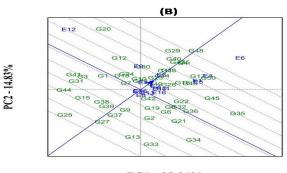
Fig 5 Ranking of gemotypes with reference to ideal genotypes

The environmental locations *viz.*, Pantnagar, Vijapur, Hisar and Junagadh had short spikes and hence they do not exert strong interactive forces.

#### 3.5. Stability analysis by GGE model

## 3.5.1. Discriminating ability and representativeness of environments

According to Yan and Thinker (2006), a long environmental vector had high discriminating ability whereas short vectors had low discrimination. It also identifies the test environment that effectively spots the superior genotypes for mega environments along with its representativeness (Yan et al. 2007). The vector length that is the absolute distance between the marker of an environment and the plot origin is a measure of the discriminating ability as the longer vector the discrimination of the environment increases. Therefore, regarding grain yield, test locations Durgapura and Jabalpur were identified as the most discriminating (informative), whereas Pune was least discriminating (Fig.4). The distance between two environments indicates their dissimilarity in discriminating ability among genotypes. If two test locations are consistently highly correlated across the years, one location can be eliminated from the analysis without losing much genotypic information (Farshadfar et al. 2013; Rakshit et al. 2014). Close relationship between the test environments may be removed from multilocation testing of cultivars as they will generate the similar information. This will help in optimal allocation of the scarce resources during multi-location trial allocation (Rakshit et al. 2012). In the bi-plot, the environments (Indore, Karnal, Kota, Pune, Dharwad, Faizabad) have environmental vectors of similar length whereas the environments (Hisar, Delhi, Ludhiana) have environmental vectors of similar length and other have environments vectors of dissimilar length. Thus most of the test environments are highly discriminating for grain yield. The earlier study (Sabaghnia et al. 2013) also



PC1 - 23.24%

Fig 6 Bi-plot for mean VS stability

reported that the length of locations vector can be used to estimates the standard deviation within each location. Observed mixture of crossover and non-crossover types of GEI in Multi- environment (MET) data is of very common occurrence (Rao *et al.* 2011) and is the indicative if existence of mega-environment within the testing locations. Discrimination ability of the location is measured by the length of the environment vectors and in the present study testing locations can be ranked from top to bottom *viz.*, Durgapura> Jabalpur > Hisar > Ludhiana = Delhi >Powarkheda>Pantnagar> Kota = Indore >Vijapur = Karnal >Dharwad = Faizabad > Pune.

#### 3.5.2. Assessment of Ideal environment :

The environment that has both high mean yield and high stability is called an ideal environment. Accordingly environments closer to the ideal environment on the biplot are regarded as more favorable than others (Farshadfar et al. 2013). The ideal environment should be most discriminative and more representative of the largest environment. The centre of the concentric circle is an ideal test environment (Fig.4). This is a point on the Average environmental axis (AEA) in the positive direction (more representative), with a distance to the biplot origin equal to the longest vector of all environments (most informative) reported by Yan and Tinker (2006). In the present study, no environment was located at the centre of the AEA therefore no environment is considered absolutely stable (Fig.4), but the Durgapura environment is to be considered a best environment because it is located near the centre of AEA. Moreover, Hisar, Delhi, Ludhiana, Pantnagar, Powarkheda, Indore, Karnal, Dharwad, Jabalpur and Vijapur are located in the above average sector. Whereas Pune, Kota and Faizabad are below average in term of grain yield and were poorest environments for selecting cultivars adapted to the whole region. Therefore, biplot for ideal environment indicated that Durgapura, Hisar, Delhi, Ludhiana, Pantnagar, Powarkheda, Indore, Karnal, Dharwad, Jabalpur and Vijapur were favorable

environments because they located in the above average sector in biplot that can be recommended testing of bread wheat genotypes.

3.5.3. Which-won-where and mega environment identification: The GGE biplot Which-Won-Where (Fig.4), which is based on a tester-centered (G + GE) table, without any scaling and it is row metric preserving. Several researchers (Yan et al. 2000; Yan and Timker, 2006; Rakshit et al. 2012; 2014) also reported that the most attractive feature of GGE biplot is which-won-where analysis in which crossover genotype  $\times$  environment interactions (GEI), mega-environment differentiation and specific adaptation of genotypes are graphically represented. The polygon (biplot of which-won-where) is formed by joining the markers of the genotypes that are farthest from the biplot origin, such that all other genotypes are placed within polygon and also a set of equality and/or perpendicular lines drawn from biplot origin to each side of the polygon. These equality and/or perpendicular lines divide the polygon into several sectors. All the genotypes in biplot were arranged in such a way that some of them were on the vertex and the rest were inside the polygon. The winning genotype for each sector is the one located at the perspective vertex (Fig.5). The genotype located at the vertices of the polygon revealed the best or the poorest in one or other environment (Yan and Tinker, 2006). These vertex genotypes were the most responsive genotypes because they have the longest distance from the origin of biplot. The responsive genotypes were those having either the best or poorest performance in one or all environments (Yan and Rajcan, 2002) falling within the sectors. In the present investigation, "Which-Won-Where" biplot divided by equality lines into seven (7) sectors with eight genotypes viz., G25, G28, G33, G34, G35, G44, and G48 as the vertex genotypes (Fig.5). All the 16 testing locations were spread in five sectors within the biplot, six in one, three each in two separate and two each in two separate sector and this may be probably due to latitudinal and longitudinal differences. The environments viz, Hisar, Delhi, Ludhiana, Dharwad, Vijapur and Karnal fall in one sector in which G35 was the winning genotype. This means that G35 was the best genotype for these above environments. The environments viz., Kota and Pune fall in the sector in which G33 was the vertex genotype; Pantnagar and Durgapur fall in the sector in which G48 was the vertex genotype; Powarkheda and Indore fall in the sector in which G29 was the vertex genotype and Faizabad fall in the sector in which G25 was the vertex genotype. No environment fall into sectors with G34 and G44 as the vertices, indicating that these genotypes were

not the best in any environments. Similar findings were reported earlier (Rakshit *et al.* 2012; 2014; Singh *et al.* 2019). The present investigation also showed that instead of conducting multi-location trials (MLTs) across closely related locations, near similar conclusion could be drown from fewer locations clustered within a mega-environment (ME). However, reported grouping of environments need to be reconfirmed using MLTs data over larger years of the data as demonstrated by Casanoves *et al.* (2005). The environments within the same sector share the same winning genotype and environment in different sector share different winning genotypes. The identification of best genotype for each location using which-won-where was earlier reported in dry bean (Mathobo and Marais, 2017).

In light of the cited logical conclusion, it can be concluded that high yielding and stable genotypes efficiently selected based on the three stability parameters bi-plot models(Eberhart and Russell model, GGE and AMMI) using multi-environmental trial (MET) data. All the testing locations showed their suitability to be used for conducting multi-location trials on the basis of their distinctness and representativeness. It was also observed that the genotype which is giving stable performance for one environment not necessarily be stable for yield in other environment. Among the tested materials, G48 ranked best for (Durgapura and Pantnagar); G29 (Powarkheda); G35 (Hisar, Delhi and Ludhiana) based on grain yield among all genotypes. Durgapura was observed as an ideal and representative environment for studying yield stability in bread wheat.

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#### 5. References

- 1. Allard RW and AD Bradshaw 1964. Implications of genotype-environmental interactions in applied plant breeding. *Crop Science* (4):503-507.
- Anonymous. 2019. Progress Report of all India Coordinated Wheat and Barley 2018-19, Crop Improvement, 1-13 pp Gyanendra Singh *et al.* (Ed). ICAR-Indian Institute of Wheat and Barley Research, Karnal.
- Becker HC and J Leon. 1988. Stability analysis in plant breeding. *Plant Breeding* 101: 1–23.

- Blanche SB, Gerald O. Myers and Kang MS (2008). GGE Biplots and Traditional Stability Measures for Interpreting Genotype by Environment Interactions. *Journal of Crop Improvement* (20):1-2, 123-135, DOI: 10.1300/J411v20n01\_07.
- Casanoves F, R Macchiavelli and M Balzarini. 2005. Error variation in multi-environment peanut trials. *Crop Science* 45: 1927–1933.
- 6. Crossa J. 1990. Statistical analysis of multi-location trials. *Advances in Agronomy* 44:55–85.
- Ebdon JS and HG Gauch. 2002a. Additive main effect and multiplicative interaction analysis of national turfgrass performance trials: I. Interpretation of genotype × environment interaction. *Crop Science* 42: 489–496.
- Eberhart SA and RA Russell. 1966. Stability parameters for comparing varieties. *Crop Science* 6:36-40.
- Farshadfar E, M Rashidi, MM Jowkar and H Zali. 2013. GGE Biplot analysis of genotype × environment interaction in chickpea genotypes. *European Journal of Experimental Biology* 3(1): 417–423.
- 10. Gauch HG. 1988. Model selection and validation for yield trials with interaction. *Biometrics* 44: 705–715.
- Kang MS. 1998. Using genotype by environment interaction for crop cultivar development. Advances in Agronomy 62: 199–246.
- Kumar A, P Kumar, G Singh and KN Tiwari. 2017. Stability analysis for different agromorphological traits under different temperature regimes in bread wheat (*Triticum aestivum L.*). *Research in Environment* and Life Science 10(3): 270–274.
- Letta T, MGD Egidio and M Abinasa. 2008. Stability analysis for quality traits in durum wheat (*Triticum durum* Desf) varieties under south Eastern Ethiopian conditions. *World Journal of Agricultural Sciences* 4(1): 53–7.
- Lin CS and Binns MR 1988. A superiority measure of cultivar performance for cultivar x location data. *Can. J. Plant Sci.* (68):193-198.
- Mathobo R and D Marias. 2017. Evaluation of genotype × environment interaction using GGE biplot on dry beans (*Phaseolus vulgaris* L.) in Limpopo province of South Africa. *Australian Journal of Crop Science* 11(5): 506–515.
- Matus-Cadiz MA, P Hucl, CE Peron and RT Tyler. 2003, Genotype × environment interaction for grain color in hard white spring wheat. *Crop Science* 43: 219–226.

- 17. Rakshit S, KN Ganapathy, SS Gomashe, A Rathore, RB Ghorade, NMV Kumar, K Ganesmurthy, SK Jain, MY Kamtar, JS Sachan, SS Ambekar, BR Ranwa, DG Kanawade, M Balusamy, D Kadam, A Sarkar, VA Tonapi and JV Patil. 2012. GGE biplot analysis to evaluate genotype, environment and their interactions in sorghum multi-location data. *Euphytica* 185:465–479.
- Rakshit S, KN Ganapathy, SS Gomashe, M Swapna, A More, SR Gadakh, RB Ghorade, ST Kajjidoni, BG Solanki, BD Biradar and Prabhakar. 2014. GGE biplot analysis of genotype × environment interaction in rabi grain sorghum [Sorghum bicolor (L.) Moench]. Indian Journal of Genetics and Plant Breeding 74(4): 558–563.
- Rao PS, PS Reddy, A Rathore, BV Reddy and S Panwar. 2011. Application GGE biplot and AMMI model to evaluate sweet sorghum *(Sorghum bicolor)* hybrids for genotype × environment interaction and season. *Indian Journal of Agricultural Sciences* 81(5):438-444.
- Sabaghnia N, R Karimizadeh and M Mohammadi. 2013. GGL biplot analysis of durum wheat (*Triticum turgidum* spp. durum) yield in multi-environment trials. *Bulgarian Journal of Agricultural Sciences* 19(4): 756–765.
- 21. SAS Institute Inc. 2011. Base SAS® 9.3 Procedures Guide. Cary, NC: SAS Institute Inc.
- 22. Shukla GK 1972. Some statistical aspects of partitioning genotype environmental components of variability. *Heredity* **29**: 237-245.
- 23. Singh C, A Gupta, V Gupta, P Kumar, R Sedhil, BS Tyagi, G Singh, R Chatrath and GP Singh 2019. Genotype x environment interaction analyses of multi- environment wheat trials in india using AMMI and GGE biplot models. *Crop Breeding and Applied Biotech* 19: 309-318.
- Singh RK and BD Chaudhary 1985. Biometrical Methods in Quantitative Genetic Analysis. (3<sup>rd</sup> Ed.), Kalyani Publishers, New Delhi, India.
- 25. Yan W and MS Kang 2003. GGE Biplot Analysis: A graphical tool for breeders, geneticists, and agronomists. CRC Press, Boca Raton, p 213.
- Yan W and NA Tinker 2006. An Biplot analysis of multi-environment trial data; Principles and applications. *Canadian Journal of Plant Sciences* 86: 623-645.
- 27. Yan W and I Rajcan 2002. Biplot analysis of test sites and trait relations of soybean in Ontario. *Crop Science* **42:** 11–20.

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- Yan W, LA Hunt, Q Sheng and Z Szlavnics 2000. Cultivar evaluation and mega-environment investigation based on the GGE biplot. *Crop Science* 40: 597–605.
- 29. Yan W, MS Kang, B Ma, S Wood and PL Cornelius 2007. GGE biplot vs. AMMI analysis of genotype-by environment data. *Crop Science* **47**: 643–655.
- 30. Yan W, PL Cornelius, J Crossa and HL Hunt 2001. Two types of GGE biplot for analyzing multienvironment trial data. *Crop Science* **41**: 656–663.
- Yau SK 1995. Regression and AMMI analyses of genotype × environment interactions: An empirical comparison. *Agronomy Journal* 87(1): 121–126.
- 32. Zobel RW, MJ Wright and HG Gauch 1988. Statistical analysis of a yield trial. *Agronomy Journal* **80:** 388–393.